

# A SURVEY OF GASTROINTESTINAL PARASITES OF CERVUS UNICOLOR (SAMBAR) IN THE KATEPURNA SANCTUARY, AKOLA

Milind Shirbhate<sup>1</sup>, Amrita Shirbhate<sup>2</sup>

<sup>1</sup>Department of Zoology, Shankarlal Khandelwal Arts, Science and Commerce College, Akola, Maharashtra, India

<sup>2</sup>Department of Zoology, Mahatma Phule Arts and Science College, Patur Dist, Akola, Maharashtra, India

## ABSTRACT

To study the prevalence of gastrointestinal parasites of Cervus unicolor, 35 faecal samples were collected from Katepurna Sanctuary during the period from February 2014 to April 2016. Twenty nine samples found be were to infected with parasites gastrointestinal and overall prevalence 82.85%. rate was Paramphistomum sp. (34.28%), Ascaris sp. (28.57%) Fasciola sp. (17.14%), stomach worm (17.14%), hook worm (14.28%), Strongyloides sp. (5.71%), Balantidium coli (5.71%) *Oesophagostomum* sp. (2.85%), Eimeria sp. (2.85%) & Mixed type of infections of Ascaris sp. Was found in 5 different scat samples or droppings.

Seasonal prevalence of gastrointestinal parasites in summer and winter were 80.01% and 17.3% respectively. This study provided a first overview on parasites in Sambar in the Katepurna Sanctuary, but to evaluate parasite transmission dynamics much more studies were required on livestock and on wild herbivores.

Keywords: *Cervus unicolor*, Fecal sample, Parasite, Scat analysis.

#### I. INTRODUCTION

The Katepurna Sanctuary in Akola, Maharashtra is an exotic sanctuary dotted with an abundance of flora and fauna. Positioned in Akola district in Vidarbha region of the state of Maharashtra, the sanctuary lies in close proximity to the catchments area of Katepurna reservoir (Mahan Dam). Its area is 20°25'0.54"N geographically located at -77°10'50.14"E. The land vegetation at Katepurna Sanctuary is southern tropical dry deciduous forest. There are over 115 species of plants at this sanctuary such as Bihada, Dhawada, Moha, Tendu, Khair, Salai, Aola, Tendu, etc. Katepurna Wildlife Sanctuary is renowned for the four-horned antelope and barking deer. Other animals that can see at the sanctuary include Sambar. Black buck. Leopard, Wolf, Wild boar, Hyaena, Hare, Nilgai, Jungle cat and Monkeys. The Katepurna water reservoir attracts many water birds.

The sambar (*Cervus unicolor*) is a large deer native to the Indian subcontinent, southern China, and Southeast Asia that is listed as Vulnerable on the IUCN Red List since 2008. Populations have declined substantially due to severe hunting, insurgency, and industrial exploitation of habitat. The name "sambar" is also sometimes used to refer to the Philippine deer, called the "Philippine sambar" and the Javan rusa, called the "Sunda sambar" (Timmins 2015). Takatsuki (1986) have studied the food habits of Sika deer (*Cervus nippon*) on Mt Goyo, Northern Honshu (The main Island of Japan) and discussed the utilization of S. nipponica as a forage for Sika deer.

#### **II. METHODS AND MATERIAL**

The material for this study comprises the faecal samples of Sambar (*Cervus unicolor*), in and around Katepurna Sanctuary and also the

agricultural land. Periods of collection of material extends from February 2014 to April 2016. A total 35 faecal samples were screened for the study of parasitic infections.

# **Collection of Faecal Material/ droppings:**

To determine the food habits, the faecal samples of Sambar (*Cervus unicolor*) were collected during study period or field visits. Faecal samples were collected in (Zip-log) polythene bags. Fresh samples were preferred for analysis. The polythene bags containing the faecal samples were labeled with date, time, locality. The bags were properly sealed and were brought to the laboratory. The size and shape of faecal pellets are also noted.



Droppings of Cervus unicolor

#### III. RESULTS AND DISCUSSION Laboratory analysis of the samples for presence of any parasitic infection:

Every time half of the faecal sample was used for observation of any parasitic infection. It was washed in warm mammalian saline and then sieved through a fine mesh. The contents were centrifuged. The supernatant was discarded and the matter settled down was observed carefully under dissecting microscope and then through compound microscope.

## **Identification of the parasite:**

The parasitic infections (Whole mounts, Eggs, Cysts) in the faecal sample were identified, separated with needle and their slides (Whole mounts) were prepared for using standard methods. Observations and identification was carried out by using standard keys and incidence was noted.

## **Evaluation of incidence :**

During screening the different samples were examined as per their habitat and incidence of parasitic infections and their percentage were noted. Total percent of parasitic forms found in *Cervus* evaluated,

overall percentage of parasitic infections of zoonotic importance.

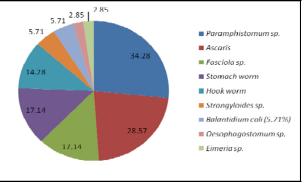
# A. Figures and Tables

#### TABLE I

SHOWING DETAILS OF PARASITES FOUND IN SAMBAR (*Cervus unicolor*) and their Incidence in %

Sr.	Name of Parasite	No	%
No			occurrence
1	Paramphistomum sp.	12	34.28
2	Ascaris	10	28.57
3	Fasciola sp.	6	17.14
4	Stomach worm	6	17.14
5	Hook worm	5	14.28
6	Strongyloides sp.	2	5.71
7	Balantidium coli (5.71%)	2	5.71
8	Oesophagostomum sp.	1	2.85
9	Eimeria sp.	1	2.85

FIG 1: SHOWING INCIDENCE IN %



IV. CONCLUSION

From the above observations it is concluded out of the 35 faecal samples Twenty nine samples were found to be infected with gastrointestinal parasites and overall prevalence was 82.85%. Paramphistomum rate sp. (34.28%), Ascaris sp. (28.57%) Fasciola sp. (17.14%), stomach worm (17.14%), hook worm (14.28%),Strongyloides sp. (5.71%),Balantidium coli (5.71%) Oesophagostomum sp. (2.85%), *Eimeria* sp. (2.85%) & Mixed type of infections of Ascaris sp. was found in 5 different scat samples or droppings. As these animals are the part of food chain in the ecosystem, further necessary treatments should be opted for the healthy survival of the species in the protected area. Our study provides a first

#### INTERNATIONAL JOURNAL OF CURRENT ENGINEERING AND SCIENTIFIC RESEARCH (IJCESR)

overview on parasites in Sambar (Cervus unicolor) in the Katepurna Sanctuary and transmission dynamics, much more studies are required on livestock in the area and on wild So, further study may also be conducted to keep assess the losses on economic point of view, due to parasitic diseases of Sambar.

## **V. REFERENCES**

- [1] Banerjee PS, Rajat G, Yadav CL and Hira R (2005). Parasite infection in some wild Animal Science 75: 206-208.
- [2] Chapman D and Chapman N (1997). Fallow deer: their history, distribution and biology: Machynllenth Coch-ybonddu Books, 139-156.
- [3] Chroust K and Chroustova E (2004). Giant [14] Mason P (1994). Parasites of deer in New liver fluke (Fascioloides magna) in cervids in South Czech regions. Veterinarstvi 54: 296-304.
- [4] Cisek A, Balicka-Ramisz A, Ramisz A and Pilarczyk B (2004). Monitoring of parasitic fauna in wild animals in the Western Pomerania Region. Folia Agriculturae Stetinensis, Zootechnica 46: 15-20.
- [5] Cook TW, Ridgeway BT, Androeid R and Hodge J (1979). Gastrointestinal helminths in diseases 15: 405-407.
- [6] Fletcher TJ (1982). Management problems and disease in farmed deer. The Veterinary Record 111: 219-223.
- [7] Fox MT (2000). Pathophysiology of nematode parasitism gastrointestinal ruminants-an update. Acta Parasitology 45: 253.
- [8] Goossens E, Dorny P, Boomker J, Vercammen F and Vercruysse J (2005). A 12month survey of the gastrointestinal helminths zoos in Belgium. Veterinary Parasitology 127(3-4): 303-312.
- [9] Islam SKMA, Ahmed S, Hoque MA, Alim [20] Parasani HR, Momin RR, Maradin MG and MA and Hassan MM (2003). Gastrointestinal parasites of captive deer and their response to selected anthelmintic. Bangladesh Veterinary Journal 37: 63-66.
- [10] Kanungo S, Das A, Das GM and Shakif-ul- [21] Pilarczyk B, Balicka-Ramisz A, Ramisz A Azam (2010). Prevalence of gastro-intestinal helminthiasis in captive deer of Bangladesh.

Wayamba Journal of Animal Science 2: 42-45.

- nearby agricultural land, but to evaluate parasite [11] Lim YAL, Ngui R, Shukri J, Rohela M and Mat NHR (2008). Intestinal parasites in various animals at a zoo in Malaysia. Veterinary Parasitology 157: 154–159.
- restores the ecological balance as well as to [12] Mackintosh CG and Wilson PR (2003). Impact of diseases on the NZ deer industry. In 63rd Conference of the New Zealand Society of Animal Production, Queenstown, New Zealand, 25-27 June, 2003. Proceeding of the New Zealand Society of Animal Production 63: 262-268.
- animals of Uttaranchal. Indian Journal of [13] Maia MJ (2001). The helminth fauna of the red deer (Cervus elaphus) and fallow deer (Dama dama) in Tapada Nacional de Mafra. Revista Portuguesa de Ciencias Veterinarias 96: 81-84.
  - Zealand. New Zealand Journal of Zoology 21: 39-47.
  - [15] Mason PC and Gladden NR (1983). Survey of internal parasitism and anthelmintic use in farmed deer. New Zealand Veterinary Journal 31: 217-220.
  - Universites [16] Mckenzie ME and Davidson WR (1989). Helminth parasites of intermingling axis deer, wild swine and domestic cattle from the island of Molokai, Hawadii. Journal of wildlife diseases 25: 252-257.
- white tailed deer of Iuinois. Journal of wildlife [17] Mutani A, Kamara R and Gabriel B (2003). А preliminary investigation on the gastrointestinal helminths of the Barbados Cercopithecus green monkey, aethiops sabaeus. Revista do Institute de Medicina tropical SaoPaulo 45: 193-195.
  - in [18] Novobilsky A, Horackova E, Hirtova L, Modry D and Koudela B (2007). The giant liver fluke Fascioloides magna (Bassi 1875) in cervids in the Czech Republic and potential of spreading to Germany. Parasitology its Research 100: 549-553.
- of antelopes, gazelles and giraffids kept at two [19] Pacon J (1994). Parasites of mouflons, stags and roe-deer from the Lower Silesia region. Wild Parasitology 40: 279-92.
  - Veer S (2001). A survey of gastrointestinal parasites of captive animals at Rajkot munipical corporation zoo, Rajkot, Gujarat. Zoo's print journal 16: 604-606.
  - and Lachowska S (2005). The occurrence of intestinal parasites of roe deer and red deer in

the Western Pomerania voivodeship. Wiadomosci Parazytologiezne 51: 307-310.

[22] Rehbein S, Walburga L and Visser M (2001). Winter Beitrage zur Kenntnis der parasitenfauna des Wildes in Nordrhein-Westfalen. Deer Endoparasitenbefall des Damwildes. Z Jagdwiss 47: 1-16.

[23] Santin DM, Alunda JM, Hoberg EP and Fuente CDL (2004). Abomasal parasites in wild sympatric cervids, red deer, Cervus elaphus and fallow deer, Dama dama, from three localities across Central and Western Spain: relationship to host density and park management. Journal of Parasitology 90: 1378-1386.

[24]Shirbhate M. V. (2007). Quantification of predation and incidence of parasitic infestation in Melghat Tiger reserve with special reference to Leopards (*Panthera pardus*). The Bioscan 2 (1): 41-46.

[25] Shirbhate M. V. (2008). Quantification of predation and incidence of parasitic infestation in Melghat Tiger reserve with special reference to Tigers (*Panthera Tigris*). The Ecoscan 2 (2): 229-235

[26] Shirbhate M. V. (2008). Quantification of predation and incidence of parasitic infestation in Melghat Tiger reserve with special reference to Wild dogs (*Cuon alpinus*). Amravati University Research Journal 3 (I): 26-32

[27] Shirbhate M. V. (2009). Feeding habits of Barking deer (*Muntiacus muntjac*) from Melghat Tiger Reserve, Maharashtra, India. The Biosphere 1 (1):71-74.